

## PATENT COOPERATION TREATY

PCT

## NOTIFICATION OF ELECTION

(PCT Rule 61.2)

From the INTERNATIONAL BUREAU

To:

Commissioner  
 US Department of Commerce  
 United States Patent and Trademark  
 Office, PCT  
 2011 South Clark Place Room  
 CP2/5C24  
 Arlington, VA 22202  
 ETATS-UNIS D'AMERIQUE  
 in its capacity as elected Office

<b>Date of mailing (day/month/year)</b> 28 June 2001 (28.06.01)	
<b>International application No.</b> PCT/EP00/09872	<b>Applicant's or agent's file reference</b> WO-02970
<b>International filing date (day/month/year)</b> 03 October 2000 (03.10.00)	<b>Priority date (day/month/year)</b> 04 October 1999 (04.10.99)
<b>Applicant</b> LANGEVELD, Pieter, Cornelis et al	

1. The designated Office is hereby notified of its election made:

☒ in the demand filed with the International Preliminary Examining Authority on:  
 02 April 2001 (02.04.01)

☐ in a notice effecting later election filed with the International Bureau on:  
 \_\_\_\_\_

2. The election ☒ was  
☐ was not

made before the expiration of 19 months from the priority date or, where Rule 32 applies, within the time limit under Rule 32.2(b).

<b>The International Bureau of WIPO</b> 34, chemin des Colombettes 1211 Geneva 20, Switzerland	<b>Authorized officer</b>  Juan Cruz
Facsimile No.: (41-22) 740.14.35	Telephone No.: (41-22) 338.83.38

# PATENT COOPERATION TREATY

# PCT

## INTERNATIONAL SEARCH REPORT

(PCT Article 18 and Rules 43 and 44)

Applicant's or agent's file reference <b>WO-02970</b>	<b>FOR FURTHER ACTION</b> see Notification of Transmittal of International Search Report (Form PCT/ISA/220) as well as, where applicable, item 5 below.	
International application No. <b>PCT/EP 00/ 09872</b>	International filing date (day/month/year) <b>03/10/2000</b>	(Earliest) Priority Date (day/month/year) <b>04/10/1999</b>
Applicant  <b>DSM N.V.</b>		

This International Search Report has been prepared by this International Searching Authority and is transmitted to the applicant according to Article 18. A copy is being transmitted to the International Bureau.

This International Search Report consists of a total of 2 sheets.  
☒ It is also accompanied by a copy of each prior art document cited in this report.

**1. Basis of the report**

- a. With regard to the **language**, the international search was carried out on the basis of the international application in the language in which it was filed, unless otherwise indicated under this item.
- ☐ the international search was carried out on the basis of a translation of the international application furnished to this Authority (Rule 23.1(b)).
- b. With regard to any **nucleotide and/or amino acid sequence** disclosed in the international application, the international search was carried out on the basis of the sequence listing :
- ☐ contained in the international application in written form.
- ☐ filed together with the international application in computer readable form.
- ☐ furnished subsequently to this Authority in written form.
- ☐ furnished subsequently to this Authority in computer readable form.
- ☐ the statement that the subsequently furnished written sequence listing does not go beyond the disclosure in the international application as filed has been furnished.
- ☐ the statement that the information recorded in computer readable form is identical to the written sequence listing has been furnished

2. ☐ **Certain claims were found unsearchable** (See Box I).

3. ☐ **Unity of invention is lacking** (see Box II).

**4. With regard to the title,**

☐ the text is approved as submitted by the applicant.

☒ the text has been established by this Authority to read as follows:

**ONE STEP TEST TO DETECT ANTIMICROBIAL RESIDUES IN EGGS**

**5. With regard to the abstract,**

☒ the text is approved as submitted by the applicant.

☐ the text has been established, according to Rule 38.2(b), by this Authority as it appears in Box III. The applicant may, within one month from the date of mailing of this international search report, submit comments to this Authority.

**6. The figure of the drawings to be published with the abstract is Figure No.**

☐ as suggested by the applicant.

☐ because the applicant failed to suggest a figure.

☐ because this figure better characterizes the invention.

☐ None of the figures.

## INTERNATIONAL SEARCH REPORT

International Application No

PCT/EP 00/09872

A. CLASSIFICATION OF SUBJECT MATTER  
IPC 7 G01N33/94 C12Q1/18

According to International Patent Classification (IPC) or to both national classification and IPC

## B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)  
IPC 7 G01N C12Q

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practical, search terms used)

WPI Data, PAJ, CHEM ABS Data, BIOSIS, MEDLINE

## C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
A	CHEMICAL ABSTRACTS, vol. 89, no. 23, 4 December 1978 (1978-12-04) Columbus, Ohio, US; abstract no. 195540, XP002132528 cited in the application abstract & J.M. INGLIS ET AL. : " Determination of streptomycin residues in eggs and stability of residues after cooking" JOURNAL OF THE ASSOCIATION OF OFFICIAL ANALYTICAL CHEMISTS, vol. 61, no. 5, 1978, pages 1098-1102, New Brunswick NJ USA -----	1-11

☐ Further documents are listed in the continuation of box C.☐ Patent family members are listed in annex.

\* Special categories of cited documents :

- \*A\* document defining the general state of the art which is not considered to be of particular relevance
- \*E\* earlier document but published on or after the international filing date
- \*L\* document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified)
- \*O\* document referring to an oral disclosure, use, exhibition or other means
- \*P\* document published prior to the international filing date but later than the priority date claimed

- \*T\* later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention
- \*X\* document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone
- \*Y\* document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art.
- \* & \* document member of the same patent family

Date of the actual completion of the international search

25 April 2001

Date of mailing of the international search report

04/05/2001

Name and mailing address of the ISA

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Authorized officer

Van Bohemen, C

## PATENT COOPERATION TREATY

## PCT

## INTERNATIONAL SEARCH REPORT

(PCT Article 18 and Rules 43 and 44)

Applicant's or agent's file reference <b>ACC/P32395</b>	<b>FOR FURTHER ACTION</b> see Notification of Transmittal of International Search Report (Form PCT/ISA/220) as well as, where applicable, item 5 below.	
International application No. <b>PCT/EP 00/ 07697</b>	International filing date (day/month/year) <b>08/08/2000</b>	(Earliest) Priority Date (day/month/year) <b>20/08/1999</b>
Applicant  <b>SMITHKLINE BEECHAM P.L.C.</b>		

This International Search Report has been prepared by this International Searching Authority and is transmitted to the applicant according to Article 18. A copy is being transmitted to the International Bureau.

This International Search Report consists of a total of 3 sheets.

☒ It is also accompanied by a copy of each prior art document cited in this report.

## 1. Basis of the report

- a. With regard to the **language**, the international search was carried out on the basis of the international application in the language in which it was filed, unless otherwise indicated under this item.

☐ the international search was carried out on the basis of a translation of the international application furnished to this Authority (Rule 23.1(b)).

- b. With regard to any **nucleotide and/or amino acid sequence** disclosed in the international application, the international search was carried out on the basis of the sequence listing :

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☐ furnished subsequently to this Authority in computer readable form.

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☐ the statement that the information recorded in computer readable form is identical to the written sequence listing has been furnished

2. ☐ **Certain claims were found unsearchable** (See Box I).

3. ☐ **Unity of invention is lacking** (see Box II).

4. With regard to the **title**,

☐ the text is approved as submitted by the applicant.

☒ the text has been established by this Authority to read as follows:

**PROCESSES FOR THE PREPARATION OF EXO- AND ENDO-INDOLOTROPANES**

5. With regard to the **abstract**,

☒ the text is approved as submitted by the applicant.

☐ the text has been established, according to Rule 38.2(b), by this Authority as it appears in Box III. The applicant may, within one month from the date of mailing of this international search report, submit comments to this Authority.

6. The figure of the **drawings** to be published with the abstract is Figure No. \_\_\_\_\_

☐ as suggested by the applicant.

☐ because the applicant failed to suggest a figure.

☐ because this figure better characterizes the invention.

☐ None of the figures.

## INTERNATIONAL SEARCH REPORT

International Application No

PCT/EP 00/07697

A. CLASSIFICATION OF SUBJECT MATTER  
IPC 7 C07D451/02

According to International Patent Classification (IPC) or to both national classification and IPC

## B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)  
IPC 7 C07D

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practical, search terms used)

EPO-Internal, CHEM ABS Data, BEILSTEIN Data

## C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
P,X	FORBES I T: "Highly stereoselective synthesis of exo and endo indolotropanes" ✓ TETRAHEDRON LETTERS,NL,ELSEVIER SCIENCE PUBLISHERS, AMSTERDAM, vol. 40, no. 52, 24 December 1999 (1999-12-24), pages 9293-9295, XP004183654 ISSN: 0040-4039 cited in the application scheme 2 and 3	1-14
P,X	WO 99 65492 A (MCDANIEL STACEY LEIGH ;LILLY CO ELI (US); AUDIA JAMES EDMUND (US);) 23 December 1999 (1999-12-23) ✓ cited in the application scheme 1 examples 1,2 --- -/--	1,3-6,9, 10

☒ Further documents are listed in the continuation of box C.

☒ Patent family members are listed in annex.

\* Special categories of cited documents:

\*A\* document defining the general state of the art which is not considered to be of particular relevance

\*E\* earlier document but published on or after the international filing date

\*L\* document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified)

\*O\* document referring to an oral disclosure, use, exhibition or other means

\*P\* document published prior to the international filing date but later than the priority date claimed

\*T\* later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention

\*X\* document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone

\*Y\* document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art.

\*G\* document member of the same patent family

Date of the actual completion of the international search

9 April 2001

Date of mailing of the international search report

04/05/2001

Name and mailing address of the ISA

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## INTERNATIONAL SEARCH REPORT

International Application No

PCT/JP 00/07697

## C.(Continuation) DOCUMENTS CONSIDERED TO BE RELEVANT

Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
P, X	WO 00 04017 A (CRAWFORTH JAMES MICHAEL ;MERCHANT KEVIN JOHN (GB); MERCK SHARP & D) 27 January 2000 (2000-01-27) cited in the application compound (VI), (VII), (VII), (IX), (X) page 14 -page 15 ---	1, 2, 11, 12
X	FRETER K: "3-CYCLOALKENYLINDOLES" JOURNAL OF ORGANIC CHEMISTRY, US, AMERICAN CHEMICAL SOCIETY. EASTON, vol. 40, no. 17, 22 August 1975 (1975-08-22), pages 2525-2529, XP000612178 ISSN: 0022-3263 page 2525; example 4N ---	1, 3-6, 9, 10
A	JUNKO NAKAO: "Trialkylmanganate-Induced Cyclization of Allyl 2-Iodophenyl Ether, N,N-Diallyl-2-iodoaniline, and alfa-Iodo Acetal" JOURNAL OF ORGANIC CHEMISTRY, 1997, pages 1910-1911, XP002165072 examples 3C, 4C -----	2

# INTERNATIONAL SEARCH REPORT

Information on patent family members

International Application No

PCT/EP 00/07697

Patent document cited in search report	Publication date	Patent family member(s)	Publication date
W0 9965492 A	23-12-1999	AU 4819099 A EP 0969005 A US 6107307 A	05-01-2000 05-01-2000 22-08-2000
W0 0004017 A	27-01-2000	AU 4637799 A	07-02-2000

(19) World Intellectual Property Organization  
International Bureau



(43) International Publication Date  
12 April 2001 (12.04.2001)

PCT

(10) International Publication Number  
**WO 01/25795 A2**

(51) International Patent Classification<sup>7</sup>: G01N 33/94

(21) International Application Number: PCT/EP00/09872

(22) International Filing Date: 3 October 2000 (03.10.2000)

(25) Filing Language: English

(26) Publication Language: English

(30) Priority Data:  
99203264.9 4 October 1999 (04.10.1999) EP

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(81) Designated States (*national*): AE, AG, AL, AM, AT, AU,  
AZ, BA, BB, BG, BR, BY, BZ, CA, CH, CN, CR, CU, CZ,  
DE, DK, DM, DZ, EE, ES, FI, GB, GD, GE, GH, GM, HR,  
HU, ID, IL, IN, IS, JP, KE, KG, KP, KR, KZ, LC, LK, LR,  
LS, LT, LU, LV, MA, MD, MG, MK, MN, MW, MX, MZ,  
NO, NZ, PL, PT, RO, RU, SD, SE, SG, SI, SK, SL, TJ, TM,  
TR, TT, TZ, UA, UG, US, UZ, VN, YU, ZA, ZW.

(84) Designated States (*regional*): ARIPO patent (GH, GM,  
KE, LS, MW, MZ, SD, SL, SZ, TZ, UG, ZW), Eurasian  
patent (AM, AZ, BY, KG, KZ, MD, RU, TJ, TM), European  
patent (AT, BE, CH, CY, DE, DK, ES, FI, FR, GB, GR, IE,  
IT, LU, MC, NL, PT, SE), OAPI patent (BF, BJ, CF, CG,  
CI, CM, GA, GN, GW, ML, MR, NE, SN, TD, TG).

**Published:**

— Without international search report and to be republished  
upon receipt of that report.

For two-letter codes and other abbreviations, refer to the "Guid-  
ance Notes on Codes and Abbreviations" appearing at the begin-  
ning of each regular issue of the PCT Gazette.

(54) Title: DETECTION OF ANTIMICROBIAL RESIDUES IN EGGS

(57) Abstract: The present invention relates to a novel method for the rapid detection of the presence or absence of antimicrobial residues in eggs. A one step test method is described in which residues of antimicrobial compounds such as antibiotics are detected while inhibiting compounds naturally present in samples obtained from eggs, which may interfere with the test, are inactivated.

WO 01/25795 A2



## DETECTION OF ANTIMICROBIAL RESIDUES IN EGGS

### Field of the invention

5           The present invention relates to a novel method for the rapid detection of the presence or absence of antimicrobial residues in eggs.

### Background of the invention

10           The presence of antimicrobial residues in food and feed is a growing concern among the consumers due to health-related problems and the increase of drug resistant bacteria. Antibiotics are not only applied as medication, but certainly in case of poultry also widely used as antimicrobial growth promoting substances. It is well known that concentrations of antimicrobial residues in eggs may be high. In most countries, such as the countries of the European Union, Canada and the  
15           United States, Maximum Residue Levels (MRL) are regulated by legislation.

          Test methods to detect antimicrobial residues in body liquids such as milk or urine such as microbial inhibition tests (e.g. agar diffusion tests) or methods making use of selective binders (e.g. antibodies or tracers) are well known. Examples of microbiological test methods have been described in GB-A-1467439, EP 0005891,  
20           DE 3613794, CA 2056581, EP 0285792 and US 5494805. These documents all deal with ready to use tests that make use of a test organism. The test organism is mostly imbedded in an agar medium, which may contain an indicator, a buffer solution, nutrients and substances to change the sensitivity for certain antimicrobial compounds in a positive or negative way.

25           Examples of suitable test organisms are strains of *Bacillus*, *Streptococcus* or *E.coli*. In general, the principle of these tests is that when antibacterial compounds are present in a sample at a concentration sufficient to inhibit the growth of the test organism the color of an acid/base or redox indicator will stay the same. However when no inhibition occurs, growth of the test organism is accompanied by the  
30           formation of acid or reduced metabolites leading to a change in the colour of the indicator.

          These test methods are suitable for the detection of antimicrobial residues in body liquids. However up to now detection of antimicrobial residues in eggs was not possible due to the presence of antimicrobial substances, such as lysozyme, which  
35           are naturally present in high concentrations in eggs. These inhibiting substances

show inhibitory activity against the test micro-organism leading to false positive results.

In case of e.g. a milk or urine sample inhibiting substances such as lysozyme or lactoferrin can be inactivated by heating the sample, e.g. at 80°C for 10 minutes (Vermunt et. al., Netherlands Milk and Dairy Journal 47: (1) 31 – 40 (1993)), or by using well known dialysis methods (van Wall, Archiv für Lebensmittelhygiene 29: (6) 235 (1978)). After this pre-treatment the liquid sample can be used for further testing simply by following the procedures of the test. In case of a Delvotest® type of test (EP 0005891) the liquid sample can be added directly to the test, after which the test is incubated.

However heating of an egg sample at temperatures sufficient to inactivate inhibiting substances of the egg, such as lysozyme, always leads to coagulation of the sample. It was believed that such samples are not suitable for further processing anymore.

Up to now after heating at a temperature sufficient to inactivate antimicrobial substances other than antimicrobial residues, the antimicrobial residues to be detected have to be extracted from the coagulated egg sample. These extraction methods not only cost a lot of time and extra handling but even worse always lead to loss of at least part of the antimicrobial residues, if present in the sample (Inglis et. al., Journal for the Association of Official Analytical Chemists 61: (5) 1098 – 1102 (1978); Katz et. al., Journal of the Association of Official Analytical Chemists 61: (5) 1103 – 1106 (1978); Janetschke et. al., Monatshefte für Veterinärmedizin 34: (21) 824 – 826 (1979); Steiner, Monatshefte für Veterinärmedizin 45: (11) 382 – 386 (1990)). This may lead to false negative results and therefore antibiotics in consumer eggs, which of course is unacceptable from a health point of view. Moreover laboratories executing studies concerning the presence or absence of antimicrobial residues in foods are limited by the time available to execute these studies. With the present time consuming methods only a very limited amount of egg samples can be examined. Further, these assays can only be executed in well-equipped laboratories and by well-educated persons, which is also a limiting factor.

It can be concluded that up to now no suitable test method for the detection of antimicrobial residues in egg samples is available. The present methods are unreliable, time consuming and may lead to both false positive and false negative results, which in turn leads to unacceptable amounts of antibiotics in the food chain and to economic losses.

### Detail d description of th invention

The present invention provides a reliable and simple to carry out, one-step test for the detection of antimicrobial residues in eggs.

5 Unexpectedly it has been found that when an egg sample is added to a test suitable for detecting antimicrobial residues and then is incubated for a sufficient time at a sufficient temperature to inactivate natural inhibiting compounds of the egg, the test can be incubated directly after heating to determine the presence or absence of antimicrobial residues.

10 It has been surprisingly found that antimicrobial residues diffuse directly from the coagulated egg sample into the test system. Thus additional extraction methods to obtain the antimicrobial residues from the coagulated egg sample are not required.

According to the invention there is thus provided a process for determining the presence of an antimicrobial residue in an egg which process comprises:

(i) contacting the sample with a test suitable for determining the presence or absence of an antimicrobial residue in the sample;

(ii) heating the contacted sample and test for a sufficient time interval to inactivate a natural inhibiting compound e.g. lysozyme present in the sample;

20 and

(iii) incubating the contacted sample and test.

By natural inhibiting compounds are meant compounds which may disturb the test and which are naturally present in the sample such as naturally inhibiting compounds, for example lysozyme. Thus by disturbing or inhibiting is meant the behaviour of the compound on parts of the test, for example the test micro organism. The invention also provides test kit for determining the presence or absence of an antimicrobial residue in a sample of an egg, which test kit comprises:

(i) a test suitable for determining the presence or absence of an antimicrobial residue in a sample; and

30 (ii) the sample,

wherein natural inhibiting compounds e.g. lysozyme present in the sample have been inactivated.

The exact time/temperature requirements depend on e.g. the condition of the sample (e.g. the starting temperature, the volume of the sample, whole egg, egg white or egg yolk); the type of test (e.g. microbial inhibition tests or assays based on selective binders (e.g. antibodies or tracers)); or the microorganism used in the test

(e.g. thermophilic or non-thermophilic *Bacillus* or *Streptomyces* species). Of course it should be taken care of that the heat treatment will not inactivate the antimicrobial residues to be detected. The heat treatment can be executed using any method known in the art, e.g. by heating in a water bath or by using an incubator as described below.

For example the test can be performed in the following way:

1. A sample of the egg is obtained by making a hole in the egg of e.g. approximately 1-2 square cm, prick the egg-yolk, place the egg with the hole down on a bottle, after the egg is empty the bottle is closed and the sample is homogenized by shaking. Alternatively of course any other method known in the art to obtain a sample of the total egg, egg white or egg yolk can be used;
2. Add a sufficient amount of the egg sample to be tested to a test using well known methods;
3. Heat the test, e.g. for approximately 10 minutes at 80° C, to inactivate the natural inhibiting substances (e.g. lysozyme), to coagulate the egg sample;
4. Incubate the test following the standard procedures of the test and read the result.

Any test suitable for determining the presence or absence of antimicrobial residues may be used in a process of test kits of the invention. Examples are described in GB-A-1467439, EP-0005891, DE-3613794, CA-2056581, EP-028579 and US 5,494,805 which are incorporated herein by reference. Suitable tests are those in which selected sensitive microorganisms are used, e.g. microbial agar diffusion tests, or tests based on selective binding of the compound to be detected. Selective binding can be achieved using the well-known antibody technology or by using specific tracers. An example of a specific tracer is the penicillin binding protein, which is used in e.g. the Delvo-X-Press® for detecting beta-lactams.

Examples of suitable microbial agar diffusion tests are tests in which species of *Bacillus*, *Streptococcus* or *E. coli* are used. Preferably thermophilic species, e.g. *Bacillus stearothermophilus* and *Streptococcus thermophilus* are used. Examples of preferred strains are *Bacillus stearothermophilus* var. *calidolactis* C953 (deposited with the Laboratory of Microbiology of the Technical University of Delft under the accession number LMD 74.1 in 1974 and with the Centraal Bureau voor Schimmelcultures (CBS), Baarn under the accession number CBS 760.83 in 1983

were the strain is available to the public) and *Streptococcus thermophilus* T101 (DSM 4022, deposited on March 3, 1987). Both strains are very sensitive to antimicrobial compounds, especially chemotherapeutics such as sulfa compounds and antibiotics such as penicillins and tetracyclines. *E.coli* strains or other suitable gram-negative bacteria can be used for the detection of e.g. quinolones.

*Bacillus stearothermophilus* var. *calidolactis* C953 and *Streptococcus thermophilus* T101 are fast growing and have the advantage that they are thermophilic. For example the optimum growth temperature of said *Bacillus* strain is from 50° to 70°C. The test organism is therefore very suitable for a test according to the invention as it is not killed by heating to inactivate the natural inhibiting compounds which may be present in the egg sample.

When the test organism is a *Bacillus* strain, it is preferably incorporated into the agar medium in the form of a spore suspension which may be prepared and incorporated into the agar medium prior to solidification by known methods (see for example GB-A-1467439). When the test organism is a *Streptococcus* strain, the bacteria are preferably incorporated into the agar medium in the form of bacterial cells which may be prepared according to known methods (see for example EP 0285792). The concentration of the test organism in the agar medium is preferably between  $10^5$  and  $10^{10}$  colony forming units per ml of agar medium.

Suitable nutrients to enable multiplication of the test organism in the absence of antimicrobial residues are for example assimilable carbon sources (e.g. lactose, glucose or dextrose), assimilable nitrogen sources (e.g. peptone) and sources of growth factors, vitamins and minerals (e.g. yeast extract).

The growth of the test microorganism can be detected using well known methods, preferably by colour change of the agar medium of the test sample. Typically a colour indicator, preferably an acid-base or a redox indicator, is used. Examples of suitable acid-base indicators include bromocresol purple and phenol red. Examples of suitable redox indicators include brilliant black, methylene blue, toluidine blue and Nile blue. Also combinations of two or more indicators can be used.

Optionally the sensitivity of the test may be altered by adding certain substances, by changing the test conditions such as pH or concentration of buffering substances or agar or by varying the ratio of the volumes of agar and egg sample. Examples of substances that may be added to the test system to change sensitivity are nucleosides such as adenosine, or antifolates such as trimethoprim, ormethoprim or tetroxoprim, which improve the sensitivity of the test organism to

sulfa compounds. Salts of oxalic acid or hydrofluoric acid may be added to improve the sensitivity to tetracyclines. Cysteine may be added to diminish the sensitivity to penicillins.

5 The amount of egg sample (whole egg, egg white or egg yolk of any species, preferably poultry) to be added to the test depends on the test system. For microbial diffusion tests typically from 0.01 to 1.0 ml, preferably from 0.05 to 0.5 ml is added to the test using well-known methods. After addition of the egg sample the test is heated to inactivate the natural antimicrobial compounds present in the sample, e.g. lysozyme, of the egg sample. Preferably the test is heated for from 2 to 20 minutes  
10 at from 70°C to 100°C, more preferably the test is heated for from 10 to 15 minutes at from 75°C to 85°C or for from 2 to 6 minutes at about 100°C. Any other time / temperature treatment, which is sufficient to inactivate the natural inhibiting compounds of the egg without inactivating the antimicrobial residues to be detected, may be used.

15 After the heat treatment the test is incubated following the instructions of the test manufacturer. The incubation time of the test is dependent on the circumstances. In case of an agar diffusion tests using *Bacillus stearothermophilus* the test is incubated in a water bath or block heater at from 60°C to 70°C, preferably at from 62°C to 65°C. Typically, results may be obtained after 1.5 to 4 hours,  
20 preferably from 2.5 to 3.5 hours. In case of tests using selective binders, such as antibodies or tracers, the results may be obtained within about 30 minutes.

Conventional microbial inhibition tests suitable for use in the present invention include the commercial products, Delvotest®, Premi®test and BR-test® (obtainable from DSM N.V. Holland) the ADM Copan®test (Copan, Italy) and the  
25 Charm®AIM test (Charm, USA). Inactivation of the natural inhibiting compounds present in the egg sample, e.g. lysozyme, is preferably achieved by heating for example for from 5 to 15 minutes at for example from 75° C to 85°C. Alternatively any other temperature / time treatment, which is sufficient to obtain said effects, can be used.

30 In a further aspect, the invention provides a test kit for carrying out the method of the invention. This test kit contain the test and is suitable to execute the method of the invention: add the egg sample, heat to inactivate the natural inhibiting compounds of the sample, incubate the test and read the results.

Examples of kits useful for the purpose of the invention are transparent  
35 tubes, single or in a set, or combined to a block of translucent material provided with

a number of holes shaped therein (incubator). The test kit may contain solidified agar medium which may be optionally buffered; a test organism (e.g. a strain of *Bacillus* or *Streptococcus*) at sufficient colony forming units; nutrients for growth of said organism; an indicator (e.g. an acid-base or redox indicator); optionally substances to change the sensitivity for certain antimicrobial compounds in a positive or negative way. All ingredients may optionally be added to the test as a separate source, for example as a tablet or paper disc.

The test kits preferably have determined sizes. This is because of the reliability of the test. In case of a test based on agar diffusion technology, preferably tubes are used. The test unit will preferably be high enough to contain an amount of agar medium and a sample corresponding to preferably a height of from 3 to 30 mm, more preferably from 5 to 15 mm. The internal cross-sectional dimension of the test units is preferably from 1 to 30 mm, more preferably from 5 to 15 mm. The test units are preferably closed air tight during storage in which conditions they may be stored for at least several months. Of course any other test unit suitable for executing the method of the invention is included in this invention.

The volume of the agar medium in the test unit is determined by the height of the test unit, the internal cross-sectional dimension of the test unit and the percentage of the volume of the test unit, which is filled with the agar medium. The volume of the agar medium is preferably from 10  $\mu$ l to 5 ml, more preferably from 100  $\mu$ l to 1 ml.

Incubators suitable to execute the heat treatments as described in this invention can be constructed in such a way that after placing the test units in the incubator, heat and incubation treatments as described above can be done. The first heat treatment to inactivate the inhibiting compounds and optionally to form solid matrix and / or to activate the spores is executed at a higher temperature, after which the incubation of the test may continue at a lower temperature. Optionally after the incubation of the test the incubator can cool down to a temperature sufficient to stop the test.

An example of such an incubator is a block heater in which test units (e.g. ampoules) can be placed. For example in case of a conventional microbial agar diffusion test using a *Bacillus stearothermophilus* strain the incubator / block heater may contain a number of holes suitable for placing the test ampoules or test plates (e.g. Delvotest® or Premi®Test) therein. After placing the ampoules or plates the incubator heats the test to a temperature of e.g. from 75°C to 85°C for e.g. from 10

to 20 minutes after which the incubator turns to a lower temperature of from 62°C to 65°C for from 1.5 to 4 hours (incubation of the test). Of course the exact time / temperature intervals depend on many factors and will differ per type of test. This invention includes all incubators capable to execute a pre-incubation at a certain temperature for a certain period of time directly followed by an incubation at a lower temperature for a certain period of time. Optionally after the incubation of the test the incubator can cool down to a temperature sufficient to stop the test.

The process described in this invention is very simple to carry out, so that persons who perform the test do not have to be specially educated or trained.

All documents mentioned in this application are herein incorporated by reference to the same extent as if each individual application or patent was specifically and individually indicated to be incorporated by reference.

### **Example 1**

#### **Preparation of a whole egg sample**

To obtain an egg sample for examination on the presence or absence of antimicrobial residues a hole of approximately 1-2 cm<sup>2</sup> was made in the egg, the egg yolk was pricked and the egg was placed with the hole down on a bottle allowing the egg white and egg yolk to drip into the bottle. After the egg was emptied, the bottle was closed and the sample was homogenized by shaking.

### **Example 2**

#### **Inactivation of natural inhibiting compounds of the egg and examining the samples on Delvotest®**

Samples of 5 eggs (duplicate), which did not contain antimicrobial residues, were obtained according to the method described in Example 1. To inactivate the natural inhibiting compounds present in the egg sample, 100 µl of each of the 5 samples was added on Delvotest® ampoules. The test was produced according to the methods described in EP 0005891 with the nutrients present in the agar. After heating for 10 minutes at 80°C in a waterbath, the ampoules were immediately placed in a waterbath at 64°C and incubated following the instructions of the producer. After 140 minutes the colour of all tests turned from purple to yellow, indicating that no antimicrobial residues were present.



Control samples were not heated at 80°C for 10 minutes, but directly placed on the ampoule. These tests remained purple for at least 4 hours.

These results clearly demonstrate that natural inhibiting compounds in the egg sample inhibited the test leading to false-positive results. When the sample was heated as described above, the activity of the natural inhibiting compounds was eliminated and no false-positive results were observed anymore.

### **Example 3**

#### **Determination of the sensitivity of the Delvotest® according to the method described in this invention using spiked samples**

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Egg samples were obtained according to the method described in Example 1. The samples were spiked by adding Penicillin G (0 and 4 ppb) or Sulphadiazine (0 and 100 ppb). The egg samples were added to Delvotest® ampoules (see Example 2) according to the method described in this invention: heated for 10 minutes at 80°C, and then immediately placed in a waterbath at 64°C and incubated following the instructions of the manufacturer. The results were read as soon as the colour turned to yellow (after 140 minutes). The samples containing no Penicillin G (0 ppb) or Sulphadiazine (0 ppb) were negative, while the samples spiked with 4 ppb Penicillin G and 100 ppb sulphadiazine remained purple (positive).

These results clearly demonstrate that the method described in this invention is suitable for detecting antimicrobial residues in egg samples.

CLAIMS

1. A process for determining the presence or absence of an antimicrobial residue in a sample of an egg which process comprises:

5 (i) contacting the sample with a test suitable for determining the presence or absence of an antimicrobial residue in the sample;

(ii) heating the contacted sample and test for a sufficient time interval to inactivate a natural inhibiting compound, for example lysozyme, present in the sample; and

10 (iii) incubating the contacted sample and test.

2. A process according to claim 1, wherein the contacted sample and test are heated to a temperature of from 70°C to 100°C.

15 3. A process according to claim 2, wherein the contacted sample and test are heated to a temperature of from 75°C to 85°C.

4. A process according to any one of claims 1 to 3, wherein the sample and test are heated for from 2 to 20 minutes.

20 5. A process according to claim 4, wherein the sample and test are heated from 10 to 15 minutes.

25 6. A process according to claim 1 wherein the test comprises a test organism, nutrients and one or more indicators present in an agar medium.

7. A process according to claim 6 whereby the degree of growth or inhibition of growth of the test organism is determined indicating the absence or presence of the antimicrobial residue.

30 8. A test kit for determining the presence or absence of an antimicrobial residue in a sample of an egg, which test kit comprises:

(i) a test suitable for determining the presence or absence of an antimicrobial residue in a sample; and

35 (ii) the sample,

wherein lysozyme present in the sample has been inactivated.

9. Use of an antimicrobial inhibition test to test an egg sample on the presence of an antimicrobial residue.

5 10. Use of an incubator for an antimicrobial residue test to inactivate lysozyme in an egg sample.

10 11. A computer program in combination with a computer, whereby the program causes the computer to operate in such a way that the computer controls the temperaure of an incubator, whereby after starting the program, the temperature will be set at a temperature for a selected inactivation time interval whereby lysozyme will be inactivated in an egg sample, whereafter the temperature is set at a temperature whereby incubation of the test sample takes place during a selected incubation time interval.

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